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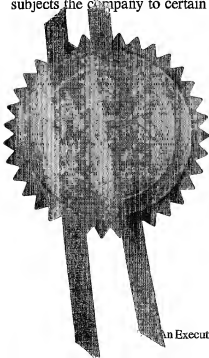WIPO   PCT

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4. Title of the invention   DNA SEQUENCES FROM AN ENDOSYMBIONT AND THEIR
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predatory protection. Many ascidians also produce large, conspicuous larvae, which develop in a brood pouch. These larvae are released in daylight hours, so they can search for optimal settlement sites, and have a short swimming phase. This strategy leaves the larvae exposed to predators and it has been suggested that selection by these predators may favour the evolution of distasteful larvae. Indeed many such larvae may be chemically defended, being unpalatable to predators when presented in feeding experiments.

There is little known of the mechanisms of production of secondary metabolites in many species. There is evidence to support the theory that symbionts (especially bacterial ones) within the invertebrate host produce at least some of these secondary metabolites, either on their own, or in conjunction with their host (Haygood, M.G., Schmidt, E.W., Davidson, S.K. & Faulkner, D.J., *J. Mol. Microbiol. Biotechnol.*, **1999**, 1, 1, 33-43: "Microbial symbionts of marine invertebrates: opportunities for microbial biotechnology"). This idea is based on the fact that some secondary metabolites show close similarities to compounds produced by bacteria. Although the evidence to support this theory is still limited, and in some cases the picture is likely to be very complex with no clear unique source for a metabolite, certain studies have added experimental support to the argument.

Although there are many secondary metabolite-producing ascidians, there are very few studies on the presence of microorganisms specifically associated with ascidians, as epi- or endobionts (Wahl, M., *J Exp. Mar. Biol. Ecol*, **1995**, 191, 239-255 : «Bacterial epibiosis on Bahamian and Pacific ascidians"; Saffo, M.B., *Marine Biology*, **1990**, 107, 291-296: "Symbiosis within a symbiosis: Intracellular bacteria in the endosymbiotic protist *Nephromyces*"). However, there are now many descriptions of symbiotic associations between bacteria and other marine invertebrate hosts. For example in sponges, molluscs, bryozoans and echinoderms.

Despite the increasing number of studies, the association between chemical defense, secondary metabolites and bacterial symbiosis, is not understood.

Ecteinascidia turbinata is the source of natural product ET-743 which is being developed as antitumoral agent. The cytotoxic compound is extracted from the ascidian which is obtained by aquaculture. Due to the life cycle of the *Ecteinascidia turbinata* this is a time consuming and laborious process.

Chemical routes to ET-743 are the total synthesis, see US,5,721,632 and the hemisynthesis from cyanosafracin B obtained by fermentation, see WO 00 69862 of the present applicant. Although these processes provide alternatives to the natural sources of ecteinascidins, they still involve numerous chemical steps and are expensive.

There is a need to provide new sources of the ecteinascidin compounds which do not have all the inconvenients mentioned before.

SUMMARY OF THE INVENTION

The present invention is directed to a DNA sequence as defined by SEQ ID 1 below or parts thereof.

SEQ ID 1:

```
ATGAATTCTGGTGGCACTGCTTAACACATGCAAGTCGAACGGTAACATAATAAATGTTTTTTACATTTATGGATGAC
GAGTGGCGGACGGGTGAGTAACGCGTAGGAACCTACCTTTTAGTGGGGGATAGCAGTGGGAAACTACTGGTAATACC
GCATGATACTTTAGAGTTAAAACTAGCTGAATTTTATAGCTTGTGCTAAAAGACGGGCCTGCGTTAGATTAGCTTGT
TGGTAAGGTAACGGCTTACCAAGGCAACGATCTATAGCTGTTCTGAGAGGAAGATCAGCCACACTGGGACTGAGATA
CGGCCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGGAAGCCTGATCCAGCAATGCCACGTG
TGTGAAGAAGGCCTTCGGGTTGTAAAGCACTTTTATTAGCGAAGAAGATATAATGGTTAAGAGCTTAATATATTTGA
CGTTAGCTAAAGAAAAAGCACCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGGAGGGTGCAAGCGTTAATCGGA
ATTATTGGGCGTAAAGAGCCTGTAGGTGGATAATTAAGTCAGATGTGAAATCCCAAAGCTTAACTTTGGAACTGCAT
TTGAAACTAATTATCTAGAGTATAGTAGAGGGTAGAGGAATTTCCGGTGTAGCGGTGAAATGCGTAGAGATCGGAAG
GAACATCAGTGGCGAAGGCGTCTACCTGGGACTAAAACTGACACTGAGAGGCGAAAGCATGGGGAGCAAACAGGATT
AGATACCCTGGTAGTCCATGCCGTAAACTATGAGTACTAACTGTTGGAATTTTTAAATTTTAGTAGTGGAGCTAACG
CAATAAGTACTCCGCCTGGGGATTACGGCCGCAAGGCTAAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCGGTG
GAGCATGTGGTTTAATTCGATGCAACGCGAAAAACCTTACCTACTCTTGAAATCCTTCGTACTTTATAGAGATATAA
AGGTGCCTTTGGAACGAAGTGACAGGTGCTGCATGGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCG
TAACGAGCGCAACCCTTTCCCTTAGTTGCCAGCGTGTAAAGACGGGGACTCTGAGGGGACTGCCGGTGATAAACCGG
AGGAAGGCGAGGACGACGTCAAGTCATCATGGTCCTTACGAGTAGGGCTACACACGTGCTACAATGGTATGTACAAA
GGGAGGCAAAATTGTAAAATCTAGCAAATCCCCAAAAGCATATCTTAGTCCGGATTGAAGTCTGCAACTCGACTTCA
TGAAGTTGGAATCGCTAGTAATCGCGAATCAGCATGTCGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGT
CACACCACGGAAGTGGAATGCACCAGAAGTGGCTAGGATAACCGAAAGGAGTCCGGTCCCTACGGTGTGTTTCGTAA
CTGGGGTGAAGTCGTAACAAGGTAGCCGTAGGGAACTGC
```

Parts of the sequence suitably have a length of at least 5, 10, 15, 20, 25, 30 or more nucleotide residues. The invention also embraces modifications or variations in the sequence SEQ ID 1, including deletions, insertions, replacements and other changes. Such modified or varied sequences typically have at least 50%, 75%, 85%, 90%, 95% or 97% homology with SEQ ID 1. References in the following description to the SEQ ID 1 or parts thereof include such modifications or variations.

In another aspect, the present invention relates to the use of the DNA sequence SEQ ID 1, or parts thereof, in an assay to identify nucleic acid molecules involved in the biosynthesis of ecteinascidin compounds, particularly by walking cromosome techniques.

The present invention is also directed to (a) DNA sequences isolated from *Ecteinascidia turbinata* in any of its development phases, its dissociated cells or cultures thereof, and/or a microorganism associated therewith, involved in the biosynthesis of ecteinascidin compounds, identified with an assay as defined above using the sequence SEQ ID 1 or parts thereof.

The invention is also directed to (b) a DNA sequence which is able to hybridize, under stringent conditions, with a DNA molecule according to (a), or parts thereof.

The invention is also directed to DNA sequences which, because of the degeneracy of the genetic code, differs from the DNA molecules according to (a) and (b) but which permits the expression of the proteins which can be correspondingly expressed using the DNA molecule according to (a) and (b), or parts thereof.

In another aspect the present invention relates to polypeptides or proteins encoded by the DNA molecules as defined above.

In another aspect the present invention is also directed to a microorganism containing a sequence according to SEQ ID 1 or parts thereof. More particularly the microorganism is a bacteria.

In another aspect, the invention is directed to the use of the microorganism as defined above in the production of an ecteinascidin compound, a precursor or intermediate thereof, or a compound involved in the biosynthesis of ecteinascidin compounds.

The invention furthermore relates to a host cell which is transformed with any one of the above-described DNA molecules.

In another aspect the invention relates to a process for the PCR amplification of a DNA molecule which is as described above.

The invention furthermore relates to a process for completing the gene cluster for biosynthesizing ecteinascidin compounds, characterized in that:
a) hybridization probes which are derived from the DNA sequence as defined by SEQ ID 1 are prepared and
b) these hybridization probes are used for the genomic screening of DNA libraries obtained from *Ecteinascidia turbinata*, and
c) the clones which are found are isolated and characterized.

DESCRIPTION OF THE FIGURES

FIGURE 1. RFLP analysis of clone inserts. HaeIII digests of 16S rDNA insert. In both cases Lane 1 = DNA marker. A. RFLP's from larval material showing the common Type 1 (lanes 2-5, 7, 9, 12), III (lane 10) and IV (lane 11) RFLP pattern. B. RFLP's from stolon material showing the Type I (lanes 4, 6-8, 10), III (lane 12), IV (lane 5) and VI (lanes 3, 9, 11) patterns. Other patterns observed are indicated by Roman numerals.

DNA SEQUENCES FROM AN ENDOSYMBIONT AND THEIR USES

FIELD OF THE INVENTION

The present invention relates to DNA sequences isolated from a bacterial endosymbiont of the ascidian *Ecteinascidia turbinata*, microorganisms containing them and their use in the production of ecteinascidin compounds.

BACKGROUND OF THE INVENTION

The ascidian, *Ecteinascidia turbinata,* is a colonial tunicate from the family Perophoridae, found in the Caribbean and Mediterranean Seas (Berrill, N.J. *Biological Bulletin,* **1932,** 62, 77-88: "Ascidians of the Bermudas"). As an important component of the benthic ecology of the Caribbean mangroves, it has been the subject of various studies examining settlement, species succession and larval behaviour , see for example:

Young, C.M. & Bingham, B.L., *Marine Biology,* **1987,** 96, 539-544: "Chemical defense and aposematic coloration in larvae of the ascidian *Ecteinascidia turbinata*"; Bingham, B.L. & Young, C.M. *J. Exp. Mar. Biol. Ecol.,* **1991,** 145, 189-204: "Larval behaviour of the ascidian *Ecteinascidia turbinata* Herdman; an insitu experimental study of the effects of swimming on dispersal"; Bingham, B.L. & Young, C.M. *Mar Ecol.,* **1995,** 16, 14S-163: "Stochastic events and dynamics of a mangrove root epifaunal community. PSZN I"; Goodbody, I., *Caribb. Mar. Stud.,* **1995,** 4, 3-10: "The timing of larval release in colonial ascidians"; Vazquez, E. & Young, *Invertebrate Biology,* **2000,** 119, 4, 433-444: "Effects of how salinity on metamorphosis in estuarine colonial ascidians".

In the 1960's interest in this species was heightened when an extract of the animal was found to have cytotoxic properties. It was not until the 1980's that the compounds conferring these properties, the ecteinascidins, were identified and characterised, see for example US 5,089,273, US 5256,663, WO9209607,

US5654,426, WO9846080. One of these compounds, ET-743, with remarkable cytotoxic acitvity, is presently in clinical trials for the treament of cancer.

10 years on, little is known about the production of these important secondary metabolites or what function, if any, they play in the animal.

Bioactive secondary metabolites are found in many marine invertebrates; especially sponges, molluscs, bryozoans and ascidians, and new compounds are constantly being described (Rinehart, K.L. **1992**, Secondary Metabolites: their function and evolution', Chadwick, D.J. & Whelan, J. (eds). Ciba Foundation Symposium 171, pp. 236-254: "Secondary metabolites from marine organisms".; Faulkner, D.J. **2000**: "Marine Pharmacology » Antonie van Leeuwenhoek 77, 135-145).

Marine invertebrate secondary metabolites encompass a wide range of chemical types including macrolides, terpenes, steroids, peptides and alkaloids, frequently with complex structures. As yet, there is little direct evidence for the function these compounds may have in their hosts, although relevant laboratory and field-based studies are beginning to address this area. Some investigations support a role in chemical defense, as anti-fouling, anti-infective or anti-predation agents (Pawlik, J.R., *Chemical Reviews*, **1993,** 93, 1911-1922: "Marine Invertebrate Chemical Defenses"). Many sponges are known to produce noxious chemicals and feeding studies have indicated that these compounds appear to confer protection against predators. Marine larvae of chemically defended adults also possess anti-predatory compounds (Lindquist, N. & Hay, M. E., *Ecology,* **1995**, 76, 4, 1347-1358: "Can small rare prey be chemically defended? The case for marine larvae").

As soft bodied, sessile marine invertebrates, adult ascidians are especially susceptible to predation, and to fouling of their external surfaces. The production of potentially defensive secondary metabolites appears widespread among certain groups of ascidians. Feeding studies using crab and fish predators have indicated that in some ascidians these metabolites confer anti-

FIGURE 2. Neighbour-joining analysis of 1288 bp from a total of 1451 bp of unambiguously aligned 16S rRNA gene sequences of Type-I (*Candidatus* Endoecteinascidia frumentensis) and IV with representative members of the gamma-Proteobacteria and an alpha-Proteobacteria outgroup. The values (>50 %) at the nodes show the bootstrap support based on neighbour-joining analysis of 100 re-sampled data sets. The scale bar represents 0.01 substitutions per nucleotide position. Phylogenetic affiliations, based on RDP II assignments, are shown to the right of the tree. GenBank accession numbers are shown alongside each representative organism.

FIGURE 3. Neighbour-joining analysis of 1289 bp from a total of 1417 bp of unambiguously aligned 16S rRNA gene sequences of Type-III and VI with representative members of the Gram positive low G+C % *Mycoplasma* and *Bacilli* as an outgroup. The values (>50 %) at the nodes show the bootstrap support based on neighbour-joining analysis of 100 re-sampled data sets. The scale bar represents 0.1 substitutions per nucleotide position. Phylogenetic affiliations, based on RDP II assignments, are shown to the right of the tree. GenBank accession numbers are shown alongside each representative organism.

FIGURE 4. Citochemical and fluorescent staining of *E. turbinata* cells. Cells of dissociated buds were stained with several dyes (A: Hemacolor. B and C: Hoechst. D: Sytox) and fixed sections of adult zooids were stained with DAPI. Samples stained with fluorochromes were visualisated under a fluorescent microscope. Fluorescent inclusions are visible with Hoechst, Sytox and DAPI (arrows). In some cases cell nucleous also appears stained (arrowheads). Bars = 10 µm.

FIGURE 5. Sections of *E. turbinata* larvae. A: A section through a whole larva stained with haematoxylin and eosin. Major anatomical features are visible: s — siphons, t — tail, b —developing branchial basket, g — gut. Cells which hybridise with probe Eub338 are often located within the developing branchial area. B to E: In situ hybridisation of larval tissue with the probe EUB338.

Fluorescent cells indicate areas of probe binding (arrows). B: A cluster of cells in the branchial basket with fluorescent inclusions. C: Three cells from a similar area to (B). D: Individual fluorescent inclusions are visible in this host cell. E: Hybridised cells from a developing larva. Bars (B — E) = 10µm. Bar (A) = 250µm

FIGURE 6. In situ hybridisation of stolon tissue with the EUB338 probe, NonEUB338 probe and EFRU-F2 specific probe (A to C) and in situ hybridisation of dissociated bud cells with the EFRU-R1 probe (D-E), using alkaline phosphatase visualisation. A dark brown / black deposit shows area of probe binding. A: Several cells show binding of the EUB338 probe (arrows). B: The NonEUB338 probe shows a very low level of background binding. C: The EFRU-F2 probe appears to be binding to a similar cell type (arrows). D: Similar cells are positive on bud tissue with EFRU-R1 probe. E: two cells showing the positive content as round-shaped elements. Bars = 10 µm.

FIGURE 7. Electron microscopy of larval tissue. A: *E. turbinata* larval cell ('bacteriocyte') showing numerous putative bacterial inclusions (arrows). N = nucleus of host cell. Bar = 2µm. B: High power image of these inclusions showing membrane detail. Bar = 1µm

DETAILED DESCRIPTION OF THE INVENTION

The present inventors directed their efforts to analyse the bacterial flora associated with *Ecteinascidia turbinata,* and identify the most commonly occurring types both in larvae and adult tissues. These bacteria could represents a potentially tractable model organism for examining this association as its secondary metabolites are well characterised and its larvae are known to be chemically defended. The presence of specifically associated bacteria, potentially an important factor in the chemical defense, could provide an alternative source of ecteinascidin compounds.

Molecular methods were employed in an effort to ensure that all associated bacteria were represented, not just those which could be grown by conventional bacterial isolation and culture techniques. In situ hybridisation analysis using probes to 16S rRNA was carried out on larval, stolon, zooid and bud tissues of *E. turbinata* in an attempt to provide localisation information, in order to confirm the specificity of the associated bacteria and to identify potentially symbiotic strains.

Surprisingly we identified rDNA corresponding to an endosymbiotic bacteria, this bacteria appears to be responsible for the biosynthesis of ecteinascidin compounds.

The 16S rDNA sequence corresponding to *Candidatus* Endoecteinascidia frumentensis SEQ ID 1 has been deposited in GeneBank with the accession number AY054370. The sequences derived from the other two strains associated with *E. turbinata* were deposited in GenBank, with the release of data withheld until the time of publication.

SEQ ID 1:

```
ATGAATTCTGGTGGCACTGCTTAACACATGCAAGTCGAACGGTAACATAATAAATGTTTTTTACATTTATGGATGAC
GAGTGGCGGACGGGTGAGTAACGCGTAGGAACCTACCTTTTAGTGGGGGATAGCAGTGGGAAACTACTGGTAAATACC
GCATGATACTTTAGAGTTAAAACTAGCTGAATTTTATAGCTTGTGCTAAAAGACGGGCCTGCGTTAGATTAGCTTGT
TGGTAAGGTAACGGCTTACCAAGGCAACGATCTATAGCTGTTCTGAGAGGAAGATCAGCCACACTGGGACTGAGATA
CGGCCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGCGGAAGCCTGATCCAGCAATGCCACGTG
TGTGAAGAAGGCCTTCGGGTTGTAAAGCACTTTTATTAGCGAAGAAGATATAATGGTTAAGAGCTTAATATATTTGA
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GAGCATGTGGTTTAATTCGATGCAACGCGAAAAACCTTACCTACTCTTGAAATCCTTCGTACTTTATAGAGATATAA
AGGTGCCTTTGGAACGAAGTGACAGGTGCTGCATGGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCG
TAACGAGCGCAACCCTTTCCCTTAGTTGCCAGCGTGTAAAGACGGGGACTCTGAGGGGACTGCCGGTGATAAACCGG
AGGAAGGCGAGGACGACGTCAAGTCATCATGGTCCTTACGAGTAGGGCTACACACGTGCTACAATGGTATGTACAAA
GGGAGGCAAAATTGTAAAATCTAGCAAATCCCCAAAAGCATATCTTAGTCCGGATTGAAGTCTGCAACTCGACTTCA
TGAAGTTGGAATCGCTAGTAATCGCGAATCAGCATGTCGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGT
CACACCACGGAAGTGGAATGCACCAGAAGTGGCTAGGATAACCGAAAGGAGTCCGGTCCCTACGGTGTGTTTCGTAA
CTGGGGTGAAGTCGTAACAAGGTAGCCGTAGGGAACTGC
```

The present invention provides one of the first reports of endosymbiotic

bacteria in an ascidian. The combination of 16S rRNA gene analysis, in situ hybridisation and electron microscopy, provides evidence for a specific association between these bacteria and the host cells. The observation of identical bacteria strains, with similar levels of dominance, and of similar cell types, in both adult (zooid and stolon) and larval tissues provide strong indications that these cells are a consistent and important feature of *Ecteinascidia turbinata*. In situ hybridisation based on the obtained sequence data suggest that the *Candidatus* Endoecteinascidia frumentensis (Type I strain) is the observed symbiont. Besides that, we found the presence of this bacterial strain in all the development stages and in samples collected all over the world, and the absence of it in a related tunicate from the same geographic area.

16S rDNA sequence analysis of dominant bacterial strains in *E. turbinata* has identified three novel strains. *Candidatus* Endoecteinascidia frumentensis falls in the gamma-proteobacteria subdivision. Although it is only distantly affiliated to any other known strains within this group, it bears similarities to species in the Legionella and Oceanospirillum subgroups. Many previously described marine endosymbionts belong to the gamma-proteobacteria subdivision, with representatives from sponges (Webster et al., 2001), bivalves (Distel et al., 1988, 1991; Sipe et al., 2000), oligochaetes (Dubilier et al., 2001) and bryozoans (Davidson & Haygood, 1997), covering a range of ecological niches and putative activities.

The spiroplasm-like Type III and VI-symbionts appear to represent a new sub group. Mycoplasms are often found as parasitic organisms in vertebrates and invertebrates, although there is increasing evidence to support both commensal and saprophytic roles. There are few records of them in marine invertebrates, although they have been identified in a bryozoan (Boyle et al., 1987). The significance of finding a Gram negative bacteria (the gamma-proteobacterium), together with two common Gram positive strains (the spiroplasms), in *E. turbinata* is not known, but it is interesting to note that a similar situation occurs in aphids (Fukatsu et al., 2000; 2001). Localisation of

the mycoplasmas in *E. turbinata* would be a valuable step forward.

The use of molecular techniques to survey bacterial populations has allowed comprehensive analysis of bacterial complement, overcoming selection for bacteria that are culturable by standard methods (Amann et al., 1995; Head et al., 1998). Use of 16S rRNA gene sequences to identify bacteria is now the standard procedure for analysis of bacteria in environmental samples and has enabled much of the current progress in the area of bacterial symbiosis. Whilst being suitable for surveying of bacterial strains, this technique can lead to misinterpretation if attempting to quantify those types present and analysis of results must take note of these possible failings. For example, Polz & Cavanaugh (1998) found that templates containing GC-rich combinations in the priming sites, might preferentially amplify.

However, the results presented below indicate such a high predominance of one novel strain, in four different tissues, that it is unlikely this could be explained by PCR bias. Similarly, bacterial types possess a different number of copies of the 16S rRNA gene, heading to potential weighting of amplification results. For example, the spiroplasm-like strains identified are likely to possess only one copy of the gene, whereas the gamma-proteobacteria strain may contain two or more. In the case of *E. turbinata* the clear predominance of strains in all tissues examined and in all the locations tested, negate the possibilities of interpretation errors.

Citochemical studies and in situ hybridisation universal probes of the examples suggests that bacteria are located within host cells in adult and larval tissue. It appears that these cells are free, possibly in the blood, and present at higher densities in the larvae. Here, they are clustered primarily in the developing branchial basket. This high density of bacteria supports the dominance observed in the 16S analysis of larval tissue, and together with in situ hybridisation using the probe to the *Candidatus* Endoecteinascidia frumentensis sequence, adds weight to the evidence that this strain is the one identified in situ.

The significance of the high numbers of 'bacteriocytes' in the branchial basket is not clear, as the larvae are not feeding, but it suggests that the bacteria may be present at sites of optimal nutrition (gaseous and nutrient exchange) as soon as the larvae has settled and the siphons have opened. However, it is also possible that a high density in this area is a side effect of the developing branchial system and that the bacteriocytes are clustered here due to blind ending blood vessels that are still growing to form the basket. Higher densities of putative bacteriocytes are also found in the stolon, which becomes engorged with cells as colonies regress. Few cell types are present, suggesting that cells have become undifferentiated, or are primarily for storage, ready to support the next phase of budding. Buds also show high density of bacteriocytes, which have probably migrated from the stolon directly. However, labeled cells are much harder to detect in zooids, where tissue is diffuse and cell density low. Still several putative bacteriocytes have been observed by in situ hybridisation, with similar characteristics as before. That is, not anchored, but freely circulating, possibly in the blood.

The forming branchial basket was the target tissue for TEM studies based on the in situ hybridisation results. The presence of bacteriocytes of similar size and apparent morphology as the cells illuminated by hybridisation in the branchial basket and in dissociated bud cells suggests that the TEM results are the same cells as those seen by hybridisation. The bacteria appear to be intracellular symbionts rather than pathogens or ingested bacteria. This is suggested from a number of observations: i) there is no obvious endocytotic behaviour by the cells; ii) the bacteria appear healthy; iii) there is no obvious pathology of the host cell in the smaller bacteriocytes; iv) the presence of so many polyribosomes suggests a large amount of synthetic activity by the cell that is intended for internal consumption rather than export; v) the bacteria appear to be of a uniform type and the same type has been observed in different individual larvae. A double membrane appears to surround the bacteria, suggesting that they are gram negative. However, the method of fixation was not optimal and further investigations of the fine structure of the

bacteriocytes may help to confirm this observation.

The presence of the bacteria in non-feeding larvae strongly suggests that they are vertically transmitted through the eggs and larvae at egg formation or during brooding. This transmission mechanism is also observed in other marine species. For example in the ascidian *Diplosoma similes* (Hirose, 2000), and in some bivalves (Krueger et al., 1996; Sipe et al., 2000). Intraovarial transmission of symbionts is also common in insects (Douglas, 1989). That the bacteria are endocytotic and likely to be vertically transmitted also suggests that they are unlikely to be culturable in simple media. Culturing experiments followed by 16S RFLP analysis of 22 bacterial strains derived from larvae, and 9 from zooids, indicate that bacteria similar to the Type I, III and VI strain did not grow under standard conditions. The difficulty in culturing symbiotic bacteria is a barrier to the further elucidation of their roles and methods to overcome this problem are needed. Schmidt et al. (2000) have managed to grow a symbiont of *Theonella* using specific culture conditions. Efforts to culture intracellular symbionts have been limited and attempts to culture host cells have generally failed (Hinde, 1971). Ways of re-addressing this problem would greatly improve the understanding of bacterial symbioses in general.

The symbionts of *Ecteinascidia turbinata* appear to be involved in the biosynthesis of ecteinascidins. Many marine symbionts are thioautotrophs from anoxic environments, oxidising sulphur and fixing CO_2. Indeed a complete sulphur cycle involving two symbionts has recently been elegantly demonstrated in a marine oligochaete (Dubilier et al., 2001). In the case of *E turbinata* it is possible that utilisation of certain nutrients in a potentially anoxic mangrove environment may be advantageous, although *E. turbinata* is known to grow and thrive in a range of habitats.

E turbinata larvae and adults are vulnerable to predation and / or fouling and production of noxious compounds may act as an effective defensive strategy. That the larvae are chemically defended, known to contain secondary metabolites and also house a large population of apparently endosymbiotic

bacteria may be circumstantial. However, it is possible that a symbiosis between the ascidian and a secondary metabolite-producing bacterium may confer their chemical protection.

Therefore, the rDNA sequence of the present invention can be used in an assay to indentify the symbiont and to isolate its DNA, for example with the widely used walking cromosome techniques. This DNA can be responsible for the biosynthesis of the ecteinascidin compounds of at least of bioprecursors or intermediates thereof. Once isolated it can be introduced in a host to provide a microorganism producing ecteinascidin compounds, providing an alternative source of these valuable compounds.

EXAMPLES

Tissue Preparation and dissection:

Ecteinascidia turbinata colonies were obtained from the Mediterranean Sea (Formentera, Spain) and Atlantic Ocean (Cádiz, Spain) and transferred to a marine aquarium at a water temperature of 24°C. Larvae were obtained from ripe colonies which were undergoing larval release, or by dissection from these colonies. They were rinsed several times in sterile filtered seawater before DNA extraction. Immature zooids (without gonads or larvae) were pinched off the colony at the stolon and incubated in sterile seawater at 24°C for 24 hours, with one change of water. They were then given a final rinse in sterile seawater and prepared for DNA extraction. Stolons from regressing colonies were removed and rinsed in sterile seawater. They were gently homogenised to liberate cells and this supernatant was removed to another tube, discarding the larger debris. The cells were spun down and pelleted, ready for DNA extraction. All material was snap frozen in liquid nitrogen for storage at —80°C. Scallop muscle *(Chlamys opercularis)* was removed from an animal held in the aquarium, rinsed in sterile seawater and used as a control.

EXAMPLE 1: 16S rDNA ANALYSIS AND RFLP's

DNA isolation, 16S rDNA PCR amplification and cloning:

All DNA manipulations were performed essentially as described by Sambrook et al., 1989. To ensure all cells, both eukaryotic and prokaryotic, were broken open, the tissue was homogenised in liquid nitrogen in a sterile mortar and pestle (except the stolon cells which were used directly). DNA was isolated either using a GNOME DNA Isolation kit (BIO101, Vista, CA), or with the following protocol. Lysis buffer (l00mM TrisHCl, l0mM EDTA, 150mM NaCl, 100□g/ml RNase A) was added to the frozen powder and allowed to thaw. The homogenised tissue was transferred to a sterile 1.5ml centrifuge tube, lysozyme was added (200pg/ml final concentration) and the sample incubated for 10 minutes at 37°C. Proteinase K and SDS were added (400 pg/ml and 1% final concentrations, respectively) and the suspension incubated for a further 30 minutes at 37°C. DNA was purified by two sequential extractions with Tris-HCl (pH 8.0) equilibrated Phenol:Chloroform:Isoamyl alcohol (25:24:1), twice with Chloroform:Isoamyl alcohol (24:1), ethanol precipitated with sodium acetate (pH 5.2) and washed with 70% ethanol. The DNA pellet was air-dried and stored in TE buffer (10 mM Tris-HCl, pH 8.0; 1 mM EDTA) at -20°C.

Two different universal 16S rDNA bacterial PCR set of primers and one set of specific oligonucleotides for DNA sequences of the *Candidatus* Endoecteinascidia frumentensis 16S rDNA were used for the amplification experiments. The universal primers were: forward, 8-AG(AG) GTT TGA TC(AC) TGG CTC AG-27; reverse 1509-G(GT)T ACC TTG TTA CGA CTT-1494 (primer position is according to *E. coli,* Weiburg et al., 1999) and 16SF1 5`-GAG A(G/C)T TTG ATC (A/C/T)TG GCT CAG-3`; 1600R 5`-AAG GAG GTG ATC CAG CC-3` (modified from Dorsch & Stackebrandt, 1992), and the specific ones were EFRU-F1, 5`-CGG TAA CAT AAT AAA TGT TTT TTA CAT TTA TG-3` and EFRU-R1, 5´-TAT GCT TTT GGG GAT TTG CTA GAT T-3` (this study). A DNA Engine (MJ Instruments, USA) thermocycler, or a Mastercycler personal (Eppendorf, Germany), cycled as follows: 94°C for 2 min., followed by 30 cycles of 55°C (30secs), 72°C (1 min 15s) and 94°C (15s), and a final elongation step of 72°C

for 10 minutes, were used. Total bacterial 16S rDNA from the extracted DNA was amplified from several dilutions to obtain optimal results and a "no-DNA" control was run for each PCR mix. The PCR product was confirmed by 1% agarose gel electrophoresis and ethidium bromide staining.

When needed, the PCR products were purified (Millipore Ultrafree-DA columns) and ligated into pGEM-T (Promega, USA) overnight at 4°C to 16°C and then transformed into competent *E. coli* DH5☐ (Life Technologies, UK). Transformants were spread on LB agar containing 50pg/ml ampicillin, 0.2 mM X-gal and 0.16 mM IPTG for blue-white screening, and incubated overnight at 37°C.

RFLP analysis of selected clones and sequencing:
Putative insert-containing clones (100 clones from each of the larval, zooid and stolon, and 40 from the scallop-control material) were selected from the *E. coli* plates by picking with a sterile toothpick and emulsifying into a 50 pl PCR reaction containing M13/pUC universal primers (forward 5'-GTT TTC CCA GTC ACG AC-3'; reverse 5'-CAG GAA ACA GCT ATG AC-3') and patched onto LB agar containing ampicillin, X-gal and IPTG (as above) for subsequent plasmid isolation. PCR reactions were cycled as follows: 94°C for 5 minutes, followed by 30 cycles of 50°C (30secs), 72°C (1min 30s), 94°C (15secs), with a final elongation step of 72°C for 10 minutes. Inserts of the appropriate size (approximately 1700 base pairs) were confirmed by 1% agarose gel electrophoresis. Positive PCR products were ethanol precipitated and resuspended in sterile dH_2O. Restriction fragment length polymorphism (RFLP) analysis was carried out using the restriction enzymes *Hae*III and *Hha*I (Promega, USA). The restricted PCR products were electrophoresed through a 1x Tris-acetate (40mM), EDTA (1mM), 3% wide-range agarose gel (Sigma, USA) and stained with ethidium bromide. Kodak 1-D system and software (Kodak, USA) was used to capture the resultant RFLP patterns. Clones representative of each different pattern observed were isolated and grown up in LB broth with ampicillin (75 pg/ml) overnight and stored frozen with 10 % glycerol at -80 °C. Plasmid DNA was isolated from representative clones for DNA sequence

analysis according to the manufacturers instructions (Qiagen Spin Mini-Preps, UK). DNA sequencing was performed using the M13 universal primers and ABI BigDye chemistry (PE Applied Biosystems, USA) and analysed on an ABI 377 DNA sequencer (PE Applied Biosystems, USA).

RESULTS:

The first approach to the analysis of the microorganisms associated to *Ecteinascidia turbinata* was done using total DNA isolated from adult zooids. Direct sequencing of the PCR amplification fragment obtained with this DNA and "universal" eubacterial primers 16SF1 (5`-GAGA(G/C)TTTGATC(A/C/T)TGGCTCAG-3`) and 1600R (5`-AAGGAGGTGATCCAGCC-3`) (modified from Dorsch & Stackebrandt, 1992) resulted in a clean sequence, indicating the prevalence of this microorganism in the tunicate. It is defined above as SEQ ID 1.

With the aim to confirm this result, specific oligonucleotides were designed for the *Candidatus* Endoecteinascidia frumentensis 16S rDNA. These oligonucleotides (EFRU-F1, 5`-CGG TAA CAT AAT A AA TGT TTT TTA CAT TTA TG-3` and EFRU-R1, 5´-TAT GCT TTT GGG GAT TTG CTA GAT T-3`) were used as primers for the PCR amplification experiments performed with total DNA isolated from adult zooids from different locations around the world (Formentera, Menorca, Túnez, Cádiz, Cuba, Florida, Puerto Rico) and with DNA obtained at different phases of development (stolon, embryos, larvae, buds and adult zooids). An amplification band with the expected size was obtained in all the cases, and RFLP and sequence analysis showed that *Candidatus* Endoecteinascidia frumentensis was present in all those samples. As a control, total DNA from a closely related organism, *Ecteinascidia conklini* was isolated, and PCR experiments were performed. No amplification was obtained with the *Candidatus* Endoecteinascidia frumentensis specific primers, although bands of the right size could be seen when universal eubacterial primers were used (data not shown). Positive results were also obtained when *E. turbinata* DNA was used (both with a sample collected at the same area as *E. conklini* was and with a sample from a different location), with universal and specific primers.

The major presence of *Candidatus* Endoecteinascidia frumentensis was confirmed by the analysis of the bacterial flora associated to the tunicate. To minimise the contribution of non-specifically associated bacteria contaminating the total DNA pool, tissue was cleaned as far as possible before DNA extraction. The incubation of live animals in sterile seawater helped to depurate the pharynx and gut of *E. turbinata,* which would have contained large numbers of non-specifically associated bacteria. Near full-length 16S rRNA gene fragments were amplified from the total DNA extracted from zooid, larval, stolon tissue and the control, scallop adductor muscle. After cloning and RFLP analysis, percentage dominance was assigned to the patterns observed in the three tissue types in order to determine any commonly occurring types. Four types were identified as occurring in all three tissue types and which also represented the most abundant RFLP types (see Table 2 and Figure 1). The Type I pattern was the most common, representing between 42-67 % of RFLP patterns observed in the three tissue types and the sequence analysis of this Type showed that it actually was *Candidatus* Endoecteinascidia frumentensis. Types VI, III and IV were the remaining three most abundant patterns (Table 2, Figure 1). The scallop control data showed no Type I (*Candidatus* Endoecteinascidia frumentensis), III or VI patterns, but a Type IV pattern was observed. This, together with the sequence analysis, identified Type IV as a commonly occurring marine-associated bacteria, and unlikely to have a specific relationship with *E turbinata.*

TABLE 2: Dominance of RFLP patterns from zooid, larval and stolon material, and the total number of patterns observed.

RFLP Type (Pattern)	% dominance of RFLP types in *E. turbinata* tissue		
	ZOOID	**LARVAE**	**STOLON**

Type	I 42.6	55.9	67.0
(*Candidatus* Endoecteinascidia frumentensis)			
Type III	5.6	9.6	5.3
Type IV	7.8	1.0	4.2
Type VI	14.6	4.3	10.0
Total number of clones digested	89	93	94
Total number of patterns observed	21	11	13

EXAMPLE 2: PHYLOGENETIC ANALYSIS

Compiled DNA sequences from two or more independent clones of each RFLP type sequenced were aligned with the RDP II database (Maidak *et al.,* 2001) using SEQUENCE MATCH and checked with CHIMERA CHECK to ensure that no sequence was chimeric. Phylogenetic inference of the novel 16S rDNA sequences was achieved by aligning the compiled 16S rDNA sequences with representative sequences obtained from the RDP II (Madiak *et al.,* 2001) and GenBank using CLUSTALX (Thompson *et al.,* 1997). Alignments were manually corrected and ambiguous positions removed. PAUP* 4.0b8 (Swofford, 2001) was used to infer phylogenetic trees using parsimony and neighbour-joining analysis according to the Kimura 2-parameter model of nucleotide substitution (Kimura, 1980) and assuming the among-site rate variation having a gamma distribution shape of 0.5. Bootstrap support for the inferred tree was established following re-sampling of 100 data sets based on neighbour-joining analysis.

RESULTS:

Four near complete 16S rRNA genes of the most commonly occurring bacterial strains were sequenced. Alignment of these sequences with the RDP II database indicated that the 16S rRNA gene sequences of Types I (*Candidatus* Endoecteinascidia frumentensis), III and VI were unrelated to any particular bacterial species in the database. The Type IV pattern was shown to have 98% sequence similarity with *Pseudomonas fluorescens* (D86001). Type I (*Candidatus* Endoecteinascidia frumentensis) could be reliably assigned as a member of the Gram negative, gamma-Proteobacteria (Figure 2). Within this group, it showed a closer relationship to the *Legionella* group. However, the deep divergence and long branch-length of this sequence makes this a tentative assignment. This was reinforced by parsimony analysis, which was not able to conclusively support the branching order as depicted in Figure 2. The closest sequence affiliation to *Candidatus* Endoecteinascidia frumentensis was an unidentified gamma-Proteobacteria (AB015255) isolated from the sediment of a deep-sea trench. Types III and VI shared 97.3 % sequence similarity to each other. Phylogenetically, they were most similar to *Spiroplasma* group, which are members of the low G+C % Gram positive *Mycoplasma* and relatives (Figure 3). Both Type III and VI, although deeply divergent from within the *Spiroplasma* group, could be reliably assigned to this group of the *Mycoplasmas*.

EXAMPLE 3: CITOCHEMICAL AND FLUORESCENT STAINING

Dissociation of buds:

Esteinascidia turbinata stolon was obtained from Cádiz (Spain) and transferred to a marine aquarium at a water temperature of 24 °C. New generations of growing buds were collected from the stolon and washed several times in sterile filtered seawater. Bud tunic was removed using forceps under sterile conditions and nude buds (20-30) were washed four times in sterile seawater. Buds were placed in dissociation medium (sterile seawater plus 0.1% collagenase), and incubated for 3 hours at 27 °C. Bud tissue was dissociated

by pipeting. To obtain a suspension of single cells, sample was left to decant on the bench for 3 minutes. The upper part of the sample was saved to other tube and cells concentrated by centrifugation at 1200 rpm/RT/4minutes. Cells were washed with 500 µl of sterile seawater, centrifuged again and resuspended in 100 µl of fixation media.

Cell fixation and slide preparation for in situ hybridisation of single cells:

E. turbinata single cells were fixed at RT for 30 minutes in 100 µl of fixation media (4% formaldehide, 5% acetic acid in sterile seawater). Fixed cells were washed with 500 µl of sterile seawater, resuspended in 20 µl of sterile seawater and spreaded on a glass slide. Slides were previously treated with TESPA in order to improve cell attachment. Slides were air dried, washed with sterile seawater and stored in 70 % ethanol (5 °C) until performing in situ hybridisation.

Cell staining

Samples of dissociated and fixated bud cells were stained with Hoechst-33342, Dapi, Sytox or Hemacolor, for 15 minutes. Cells were centrifuged (1200 rpm/RT/4 minutes), resuspended in 250 µl of sterile seawater and observed under UV light in a inverted fluorescent microscope (Leica DM IRB).

RESULTS:

Dissociated bud cells or fixed sections of adults were used for several staining experiments in order to identify intracellular microorganisms in *E. turbinata* cells. By using Hemacolor, a general staining method, intracellular positive granules could be observed in some cells of dissociated buds (Figure 4A). Several fluorescent dyes specific for nucleic acids were also employed. These dyes are sensitive enough to allow detection of DNA/RNA containing particles as bacteria and mycoplasms, but not subcellular organelles as mitochondria or chloroplasts. Hoechst and Sytox showed intracellular positive elements in cells of dissociated buds. In some cases, just a few fluorescent round or rod-shaped

elements were observed inside large vacuoles (Figure 4B) but also dense stained cells showing a granular appearance could be detected (Figure 4C and D). In fixed sections of *E. turbinata* zooids, DAPI staining showed the same type of fully stained cells (Figure 4E). The cell nucleus is visible in some cases (Figure 4A, B and E), when not hidden by fluorescent particles (Figure 4C and D).

EXAMPLE 4: IN SITU HIBRIDISATION WITH A UNIVERSAL PROBE:

In situ hybridisation was carried out using standard methods (Manz et al., 1992; Amann et al., 1990; Wilkinson, 1994). Tissue was fixed in 4% Paraformaldehyde (PFA) in TBS (0.lMTris-HCI, 0.9%NaCI) with 0.1M MOPS at room temperature for 30mins to 1 hour. It was then embedded in paraffin wax, sectioned, and sections were mounted on gelatin-coated slides. Two panels of sections were mounted on each slide, placing alternate sections on each panel, to provide a test and control panel of the same area of the tissue. The slides were then prepared for hybridisation. In brief, after rehydration they were rinsed, incubated in l mug/ml Proteinase K, post-fixed in 4% PFA in TBS for 10 minutes, rinsed and treated with 0.25% acetic anhydride in 0.1M Triethanolamine-HCI pH8, dehydrated and air dried, ready for storage at -80°C.

Initial in situ hybridisation studies were carried out using a biotinylated universal bacterial 16S rRNA probe to identify sites of potential interest (EUB338 5'-GCT GCC TCC CGT AGG AGT-3', and a control probe NON-EUB338 5'- ACT CCT ACG GGA GGC AGC-3') (Amann et al., 1990). Other controls consisted of incubations with no probe, and panels of attached bacterial cells (*E. coli*) which had been fixed and processed as the sections. Hybridisation was carried out at 45°C for 3 hours in buffer (0.9M NaCl, 20mM Tris-HCI pH7.2, lx Denhardts, 0.1% SDS, 5mM EDTA, 0.lmg/ml Poly(A)) with 2.5ng/mul of probe which had been reconstituted in TE. Hybridisation chambers with dual ports were used to prevent evaporation (Grace BioLabs:

22mm x 22mm chamber for each panel of sections on the slide). Following hybridisation the slides were washed 2 x 15 minutes in wash buffer (0.9M NaCl, 20mM Tris-HCI pH 7.2, 0.1% SDS) at 48°C. Binding of probe was visualised using Avidin-DN (Vector Labs) as recommended by the manufacturer. Sections were mounted in Vectorshield (Vector Labs) and viewed on a Zeiss Axioskop with fluorescent attachments. For visualisation with alkaline phosphatase (AP) the slides were washed after hybridisation and incubated in AP conjugated anti-biotin antibody (Vector Labs) diluted 1:1000 in buffer (20mM sodium phosphate, 0.9% NaCI, 0.1% Tween, 0.1% BSA) at 4°C overnight. They were then washed and visualised using a BCIP/NBT — AP substrate kit (Vector Labs).

RESULTS:

Hybridisation of the universal bacterial EUB 338 16S rRNA probe was observed in larval sections (Figure 5), stolon material and at low levels in zooid sections. Positive *in situ* hybridisation was found in cells of approximately 10-12 mum diameter which demonstrate a granular appearance. These cells usually had an unstained area at one pole, which was thought to be the host cell nucleus. In larvae the cells were predominantly found in the developing branchial basket (Figure 5). In zooid tissue, only a few positively hybridising cells were observed. These cells were located in the region of the pharynx and with cells possibly associated with blood vessels. The background fluorescence was relatively high, but hybridisation positive cells were easily distinguishable on the test sections, but were clearly not present in the controls. Dissected portions of regressing stolon and budding tissue were also examined by fluorescent in situ hybridisation. With these preparations, the background fluorescence under the FITC filter was too high to facilitate accurate interpretation of hybridisation. Consequently, subsequent hybridisation experiments used the histochemical AP visualisation system. Hybridising cells were found scattered throughout the stolon and bud tissue. The appearance of the hybridisation-positive cells in the stolon and bud tissue had the same general appearance as observed in the larvae, with the hybridised part of the cell exhibiting granular staining and an unstained, approximately polar region, presumably representing the host

nucleus (Figure 6A). The non–Eub338 probe gave very low levels of background binding using the AP system, such that it was almost indiscernible when compared to the intensity of Eub338 binding (Figure 6B). In all cases where the control was buffer without probe, the cells hybridised in the test sections could not be observed in the control sections.

EXAMPLE 5: SPECIFIC PROBES : DOT BLOTS AND IN SITU HYBRIDISATION:

Probes were designed to putatively unique regions of the Candidatus Endoecteinascidia frumentensis and Type III 16S rDNA sequences:

Oligonucleotide	Sequences (5`-3`)	Application[a]	References
Symbiont specific			
EFRU-F1	CGG TAA CAT AAT AAA TGT TTT TTA CAT TTA TG	P, I, S	Herein
EFRU-R1	TAT GCT TTT GGG GAT TTG CTA GAT T	I, D, S	Herein
EFRU-R2	CTT TCG GTT ATC CTA GCC AC	I, D	Herein
Domain bacteria			
Type III-1	GCA ACT ATT TCT AGC TGT TAT TC	D	Herein
Type III-4	AGC TTT GCA CTG GAT GTC AAG	D	Herein
EUB338	GCT GCC TCC CGT AGG AGT	I, D	Amann et al., 1990
NON-EUB338	ACT CCT ACG GGA GGC AGC	I	Amann et al., 1990
EUB8-f	AG(AG) GTT TGA TC(AC) TGG CTC AG	P, S	Weiburg et al., 1999
EUB1509-r	G(GT)T ACC TTG TTA CGA CTT	P, S	Weiburg et al., 1999
16S-F1	GAG A(G/C)T TTG ATC (A/C/T)TG GCT CAG	P, S	Modified from Dorsch & Stackebrandt, 1992
1600R	AAG GAG GTG ATC CAG CC	P, S	Modified from Dorsch & Stackebrandt, 1992
Universal primers			
M13/pUC-f	GTT TTC CCA GTC ACG AC	S	
M13/pUC-r	CAG GAA ACA GCT ATG AC	S	

ᵃ P= PCR primer; I= In situ hybridization probe; D= Dot-blot probe; S= Sequencing primers

Probe specificity was initially examined by submitting the probe sequence to the SEQUENCE MATCH and PROBE MATCH on the RDP II database (Maidak *et al.,* 2001) and compared to *E. coli* 16S secondary structure to check for optimal regions of binding (Amann et al., 1995; Zheng, et al., 1996).

Conditions for probe binding were confirmed by dot blot analysis. 16S rDNA PCR products from the Type I Candidatus Endoecteinascidia frumentensis, III, IV & VI plasmid clones (50ng) and *E. coli* DH5 alpha were spotted onto a positively charged nylon membrane as per manufacturers instructions (Schleicher & Schuell). The membranes were prehybridised at 46°C for 1 hour in hybridisation buffer (as previously plus 2% Marvel), and then hybridised at 46°C for 3 hours with 0.5ng/mul of probe. Membranes were washed 2 x 15 minutes at 50°C (wash buffer as before), blocked in NaPBS (20mM sodium phosphate, 0.9% NaCl) with 1% BSA for 1 hour and visualised using anti-biotin alkaline phosphatase (1:2000 in NaPBS, 0.1% BSA) and the BCIP/NBT — AP substrate kit (Vector Labs).

Following dot blots, hybridisation with EFRU-F2 probe was carried out on sections at 46°C for 3 hours, in hybridisation buffer, containing 0, 5, and 10% formamide, plus a no probe control of each, in an attempt to optimise the stringency for each probe. 2 x 15 minute washes were carried out at 50°C. Slides with control bacteria *(E. coli)* were also used at each of the formamide concentrations, with each probe, to further check for stringency. Stolon tissue was screened on sections as this provided a more uniform basis for comparisons. Successful hybridisations were repeated on larval sections.

RESULTS:

The dot blots indicated EFRU-F2 was specific for the Type I (*Candidatus* Endoecteinascidia frumentensis) sequence, binding only to Type I 16S rDNA

and not to that of the other common types isolated from *E. turbinata* or to *E. coli* 16S rDNA, under standard hybridisation conditions see Table 3.

Same results were obtained for EFRU-R1 probe.

TABLE 3: Dot-blot hybridisation of unique 16S rRNA in situ probes

	Probes			
16S rDNA PCR product	Eub338	EFRU-F2	Type III-1	Type III-4
Type I	X	X	X	-
Type III	X	-	X	X
Type IV	X	-	-	-
Type VI	X	-	X	X
E. coli	X	-	-	-
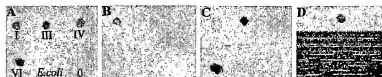
Type III-4 was also specific for Type III and VI sequence, but type III-I cross-reacted with the Type I DNA under these conditions. The Type I probe tested on stolon sections bound to similar cells as those highlighted by the universal eubacterial probe EUB338, suggesting that these bacteria are the *Candidatus* Endoecteinascidia frumentensis symbiont (Figure 6C).

In situ hybridisation of EFRU-R1 probe on dissociated bud cells showed the granular cell type previously described (Figura 6D) together with some other cells where isolated positive elements could be observed (Figura 6E). These figures could correspond to lightly crushed granular cells showing the cellular content. The round shaped elements are suggested to be putative Type I bacteria (*Candidatus* Endoecteinascidia frumentensis), which appear tightly packed in certain cell types.

EXAMPLE 6: ELECTRON MICROSCOPY:

Electron microscopy was carried out on mature larvae. Larvae were fixed as before, post-fixed in 1% Osmium tetroxide and embedded in Spurrs resin. The blocks were orientated so that sections were cut from the developing branchial basket area (an area potentially containing the cells of interest, as identified by *in situ* hybridisation) and sectioned using an LKB ultramicrotome. Sections were mounted on grids and stained using uranyl acetate and lead citrate. Observations were made using a Jeol I00S Transmission Electron Microscope.

RESULTS:

The rods of the developing branchial basket in E *turbinata* larvae were formed by elongate support cells. These were frequently vacuolated with large, electron dense inclusions (possibly lipids). Putative cells which correlated with those observed by in situ hybridisation were identified and termed "bacteriocites" (Figure 7). They had a very similar morphology to the other major type of coelomocytes except for the presence of putative bacteria. The bacteriocytes frequently had cytoplasmic protrusions extending from the cell surface and all appeared to be free in the coelom. In small cells the nuclei were heterochromatic and unremarkable. Mitochondria were frequently observed. There were some obvious strands of rough endoplasmic reticulum, but the cells were most notable for the high densities of what are apparently polyribosomes distributed throughout the cytoplasm. There was no obvious golgi apparatus.

The putative bacteria were rounded and approximately 1-2 microns in diameter. Their membrane structure suggests that they are gram-negative bacteria and they contain no obvious organelles (Figure 7). The chromosomal DNA is usually well distributed and only partially condensed. The bacteria appear to be healthy, particularly in the smaller cells, with any obvious shrinkage probably the results of the fixation technique rather than any pathology. No bacteria were seen being lysed.

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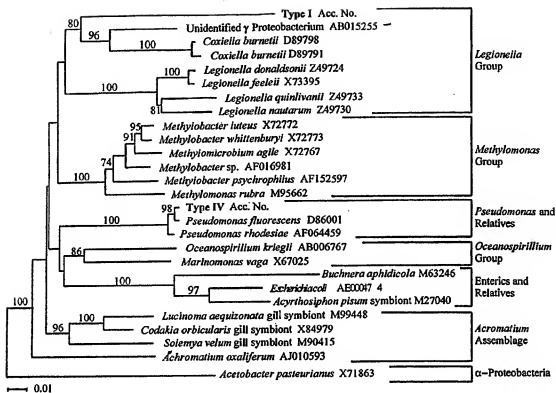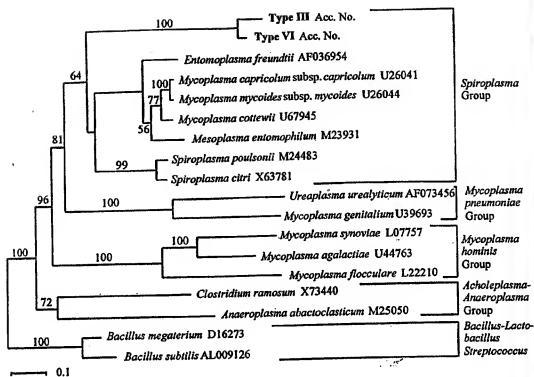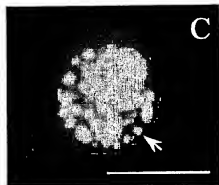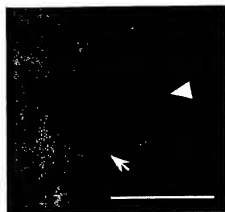
FIGURA 1

FIGURE 2

FIGURA 2

Type III Acc. No.
Type VI Acc. No.
Entomoplasma freundtii AF036954
Mycoplasma capricolum subsp. capricolum U26041
Mycoplasma mycoides subsp. mycoides U26044
Mycoplasma cottewii U67945
Mesoplasma entomophilum M23931
Spiroplasma poulsonii M24483
Spiroplasma citri X63781
Ureaplasma urealyticum AF073456
Mycoplasma genitalium U39693
Mycoplasma synoviae L07757
Mycoplasma agalactiae U44763
Mycoplasma flocculare L22210
Clostridium ramosum X73440
Anaeroplasma abactoclasticum M25050
Bacillus megaterium D16273
Bacillus subtilis AL009126

Spiroplasma
Group

Mycoplasma pneumoniae Group

Mycoplasma hominis Group

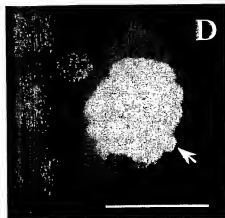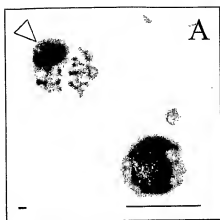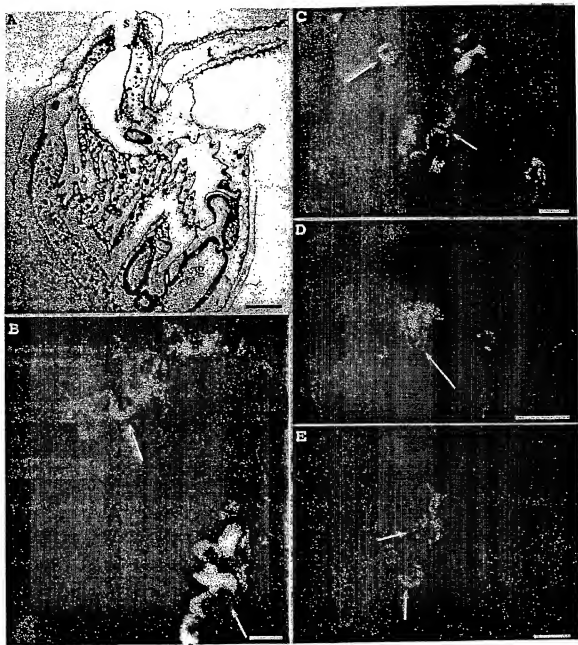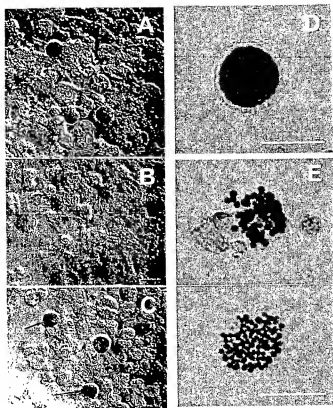*Acholeplasma-Anaeroplasma* Group

Bacillus-Lacto-bacillus Streptococcus

0.1

FIGURE 3

FIGURA 3

Figure 4

FIGURA 5

Figure 6

FIGURA 7